# Thyroid Hormone Reference Interval- Evidence from Alappuzha District, Kerala State, India

MIRIAM VARKEY<sup>1</sup>, GILSA ENGOOR SATHIANDRANATHAN<sup>2</sup>, DEEPA MARIA KUTTIKADAN VARGHESE<sup>3</sup>, SATHEESAN RAMAKRISHNAN<sup>4</sup>

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## **ABSTRACT**

**Biochemistry Section** 

**Introduction:** Thyroid diseases are one of the most common endocrine disorders not only in India, but worldwide. Most commonly done biochemical tests to diagnose thyroid disorders are Thyrotropin (TSH), thyroid hormones {free thyroxine (FT4) and free triiodothyronine (FT3)} in serum. The Reference Interval (RI) is influenced by a variety of factors like diet, genetics, iodine nutritional status and thyroid autoimmunity. Hence, the reference values of thyroid function tests commonly used in the clinical laboratories are derived from data from the western population and may not be applicable for Indian population.

**Aim:** To establish RI for thyroid hormones in the coastal area of Alappuzha District in Kerala.

**Materials and Methods:** This community-based study included 228 adults in the age group 18-80 years. TSH, FT3, FT4, Thyroperoxidase antibody (TPOAb) and anti-thyroglobulin antibodies (TgAb) were analysed on an automated immunoassay system Access 2 Beckman Coulter using a direct chemiluminescence detection system. RIs were calculated according to International Federation of Clinical Chemistry (IFCC) recommendation using IBM SPSS version 20.0 for windows.

**Results:** Rls for serum TSH, FT4 and FT3 were calculated as 0.11-6.39 mIU/L, 0.58-4.66 ng/dL and 0.72-4.66 pg/mL. No significant age and gender difference in the Rls are noted.

**Conclusion:** In this study, RI established in the local reference population found to be different from those reported by previous studies conducted in other geographical areas.

## INTRODUCTION

About 42 million people in India suffer from thyroid diseases [1]. An early diagnosis and treatment warrants better management and outcome. Biochemical tests to assess and to diagnose thyroid disorders include TSH, T3, FT3, T4 and FT4, thyroglobulin [Tg]; Thyroperoxidase antibody (TPOAb), TSH receptor antibodies (TRAb) and anti-Tg antibodies (TgAb). Measurement of TSH and thyroid hormones, FT4 and FT3 in serum is routinely done in most clinical biochemical laboratories [2]. During the last 50 years, laboratory evaluation of thyroid dysfunction has changed tremendously. Sensitivity and specificity of biochemical thyroid tests has improved greatly. The fifth generation assay systems have a sensitivity as low as <0.004 mIU/L [3]. This has strongly influenced the clinical strategy for detecting and treating thyroid diseases.

TSH is the most sensitive marker for thyroid dysfunction especially for subclinical hypothyroidism. It also has an essential role in treating both hyperthyroidism and hypothyroidism for dose adjustment. Besides, it has a prognostic significance for tumor recurrence [4]. Clinical signs and symptoms of hyperthyroidism or hypothyroidism are often non-specific and vague. Hence, estimation of thyroid hormones (total and free thyroxine, T4 and FT4; total and free triiodothyronine) is also important in the evaluation in thyroid dysfunction. For the diagnosis of autoimmune thyroid disease, TPOAb and anti-Tg are used. RI aids the clinician in interpreting observed values. The reference values of TFT used in clinical laboratories have been adopted from those reported for the western population. These RIs may not be applicable for Indian population since a variety of factors like diet; genetics and thyroid autoimmunity influence the thyroid hormone levels [5].

RIs for TFT are also influenced by iodine status. Iodine nutritional status either deficiency or excess, has a significant impact on the determination of RI of thyroid hormones. The programme of Universal Salt Iodisation (USI) programme was instituted in India in 1984 and

**Keywords:** Free thyroxin, Free triodothyronin, Thyroid disorders

various studies conducted in many parts of the country has reported iodine sufficiency [6,7]. RIs are also dependent on analytical quality parameters such as sensitivity, specificity, precision, and accuracy of the applied assay system.

RI may be established by either direct or an indirect method. In direct method, reference individuals are selected based on precisely defined criteria from a reference population according to Clinical and Laboratory Standards Institute/International Federation of Clinical Chemistry and Laboratory Medicine (CLSI/IFCC) recommendation [8]. In indirect method, instead of reference individuals, values with the required characteristics are selected from an existing database. Direct method is time-consuming and expensive and the most important step is the selection of the reference group and the standardisation of pre-analytical factors [9,10].

The studies conducted in Delhi and in Ranchi report a wider range for T4 and TSH [11,12] compared to that reported in another study from Ranchi [13]. Even though similar studies for establishment of RI was done in other parts of country, data from Kerala is found to be lacking. Thus, the aim of this study was to establish RI in a sample population from Alappuzha district in Kerala.

## MATERIALS AND METHODS

This community-based cross-sectional study was conducted in selected wards of Ambalaphuzha North Grama Panchayath of Alappuzha District in Kerala State, India from May 2017-August 2017. The study was approved (B3/1573(A)2010/TDMCA) by the local ethical committee.

According to CLSI C28-A guideline, 153 reference values were needed to establish a non-parametric RI with 99% confidence interval [14]. A total of 438 subjects were recruited to the study, after taking informed consents. A questionnaire was prepared according to the CLSI C28-A standard, for determining exclusion criteria and pre-analytic factors.

**Inclusion criteria:** Subjects from both sex in the age group of 18-80 years were included in the study.

**Exclusion criteria:** Subjects with history of thyroid disease, family history of thyroid disease, use of medications known to interfere thyroid function, visible palpable goiter and systemic illness were excluded from the study [15].

Thorough history and physical examination was done for each subject. As per recommendations from the National Health and Nutrition Examination Survey (NHANES) [14], the National Academy of Clinical Biochemistry (NACB) only euthyroid healthy volunteers were included, who were free from detectable autoantibodies against TPOAb or TgAb [15,16]. Hence, final decision about a reference person was made with the help of laboratory findings.

Two wards, ward number 2 and 18 from Ambalaphuzha North Grama panchayath were selected. Subjects according to the inclusion criteria were identified in each house of these wards with the help of ASHA workers. Instruction was given to the subjects to come for blood collection on specified date to the locally arranged blood collection camp at sub center of Ambalaphuzha primary health center and panchayat auditorium.

Blood samples were collected from 7.00 to 9.00 am, after an overnight fasting, into vacutainer tubes. Centrifugation was performed within one hour of sample collection at 1500×g for 10 minutes and samples were analysed immediately. The measurements were made using the same reagents and the same instruments for all patients. Analyses of TSH, FT3, FT4, TPOAb, and TgAb were performed in an automated immunoassay system, Access 2 Beckman Coulter using a direct chemiluminescence detection system according to the manufacturer's instructions.

TPOAb and TgAb levels above the 9 and 4 IU/mL, respectively were regarded as positive according to the manufacturer's data. Assay imprecision was assessed by the use of commercial quality-control materials, Liquicheck levels 1-3 (Bio-Rad). Each control material was analysed in duplicate per run. Daily runs were performed for five days in a week over three weeks for a total of 30 replicates for each control.

#### STATISTICAL ANALYSIS

Data was analysed using IBM Statistical Package for the Social Sciences (SPSS) version 20.0 for Windows. Outliers was identified by Tukey's method [17,18].Visual inspection of histogram, Q-Q plot, box plot and normality check using Shapiro-Wilkis test, it was found that FT3 and FT4 levels in the reference population was normally distributed, but TSH levels are having non-gaussian distribution even after log transformation of the data. Hence, according to IFCC recommendation, non-parametric methods were used for the analysis of RIs were expressed as median and 95% confidence interval. The levels of hormones were compared between males and females and among different age group by Mann-Whitney U test and by Kruskel-Wallis H test, respectively with a significant level for p-value as <0.05.

## RESULTS

Out of 446 subjects, 129 were excluded due to the increased Anti-TPO and Anti-Tg levels. From the remaining 317 subjects, data of 228 subjects were included for analysis after removal of outliers and excluding the data with other abnormal laboratory results like high blood glucose, abnormal liver and renal function tests and high lipid profile. There were 62 males and 166 females in the study population. The mean age was 43±12.35 years. The minimum age was 18 years and maximum age was 71 years [Table/Fig-1].

The levels of FT3, FT4 and TSH in the reference population are given in [Table/Fig-2]. None of the parameters showed Gaussian distribution even after the log transformation. Hence, non-parametric methods were used to establish the RI. The RI for sample population was determined as 2.5 percentile as lower limit and 97.5 percentile as

Age group	Male	Female	Total			
Group 1: 18-30 years	12	30	42			
Group 2: 31-45 years	20	58	78			
Group 3: >45 years	30	78	108			
[Table/Fig-1]: Age and gender distribution.						

upperlimit. A 90% confidence interval for both values was calculated by bootstrap method.

Parameter	N	Lower limit- 2.5 percentile (90% Cl)	Non parametric median (IQR)	Upper limit- 97.5 percentile	Manufacture's RI	
FT3 (pg/mL)	228	0.72 (0.54-1.23)	3.11 (0.92)	4.76 (4.40-4.55)	2.5-3.9	
FT4 (ng/dL)	217	0.58 (0.55-0.62)	0.83 (0.19)	4.66 (3.59-6.03)	0.6-1.1	
TSH (mIU/L)	217	0.11 (0.21-0.37)	1.65 (1.94)	6.39 (5.00-7.37)	0.34-5.2	
<b>[Table/Fig-2]:</b> Reference Interval (RI) for thyroid function tests established in the present study. IQR: Interquartile range; CI: Confidence interval; TSH: Thyrotropin; FT4: Free thyroxine; FT3: Free triiodothyronine						

Few samples could not be analysed due to inadequate volume.

The levels of FT3, FT4 and TSH in males and females were compared by Mann-Whitney U test [Table/Fig-3]. No significant difference were observed for FT3 {U(Nmale=99.82, Nfemale=109.52)=4037, Z=1.027, p=0.304}, FT4 {U(Nmale=0.931, Nfemale=109.23}=6504, Z=0.087, p=0.931).

Parameter	Gender	Ν	Mean rank	U	Z	p-value
FT3 (pg/mL)	Male Female	69 159	99.82 109.52	4037	1.027	0.304
FT4 (ng/dL)	Male Female	60 157	108.4 109.23	6504	0.087	0.931
TSH (mIU/L)	Male Female	60 157	90.53 107.76	5704	1.883	0.060
<b>[Table/Fig-3]:</b> Thyroid parameters based on gender. U: U test statistics (sum of mean ranks); Z: Z score (Standardised test statistics); TSH: Thyrotropin; FT4: Free thyroxine; FT3: Free triiodothyronine						

Krusker Wallis test showed no significant difference among different age group [Table/Fig-4]. The tests results for FT3 shows X2 (2)=3.56 and p=0.168 with mean rank score of 96.84,11.85 and 103.76, for FT4, X2 (2)=3.067 and p=0.216 with mean rank score of 122.04,100.93 and 11.07 and for TSH X2 (2)=4.662 and p=0.097 with mean rank score of 99.76,103.68 and 92.51 in age group 1,2 and 3, respectively.

Parameter	Age group (Year)	N	Mean rank	Chi-square	p-value
FT3 (pg/mL)	18-30	38	96.84	3.563	0.168
	31-45	78	11.85		
	46-71	108	103.76		
FT4 (ng/dL)	18-30	40	122.04		0.216
	31-45	76	100.93	3.067	
	46-71	101	11.07		
TSH (mIU/L)	18-30	37	99.76		0.097
	31-45	78	103.68	4.662	
	46-71	102	92.51		
<b>[Table/Fig-4]:</b> Thyroid parameters in different age group. TSH: Thyrotropin; FT4: Free thyroxine; FT3: Free triiodothyronine					

## DISCUSSION

The present study established a new RI for thyroid function test in a local reference population of Kerala, India. The new RI differs from that reported by the manufacturer. Even though the RI of TFT in adults are well-established, different studies report different values for FT4, FT3 and TSH which can be attributed mostly to the difference in assay techniques and reference population. RI reported by various investigators from other part of India and other countries are summarised in [Table/Fig-5] [11-13,19-25].

india 2013 , India 2017 dia 2015 dia 2018	ECLIA Roche CLIA Abbot CLIA Abbot ECLIA Roche	1916 153 3346 1000	1.56-5.72 1.18-3.79 1.81-3.53	0.66-1.62 0.54-1.48 0.76-1.34	0.68-9.78 0.80-9.78 0.08-4.36
lia 2015	Abbot CLIA Abbot ECLIA	3346			
	Abbot ECLIA		1.81-3.53	0.76-1.34	0.08-4.36
dia 2018		1000			
					0.27-5.63
2016	CLIA Siemens	717	2.34-3.73	0.72-1.33	0.43-5.51
2005	ELIZA	3915	2.47-4.56	0.54-1.23	0.25-2.12
2018	RIA	390			0.50-3.1
2017	ECLIA Roche	250	2.66-4.12	0.81-1.31	0.65-5.39
2014	RIA	852	1.81-3.51	0.78-1.45	
2010	CLIA Beckman	619	2.62-3.84	0.61-1.06	0.41-4.25
ia 2020	CLIA Beckman	228	0.72-4.76	0.58-4.66	0.11-6.39
-	2010 ia 2020	ia 2020 CLIA Beckman CLIA Beckman	2010 CLIA 619 Beckman CLIA 228	2010 CLIA Beckman 619 2.62-3.84   ia 2020 CLIA Beckman 228 0.72-4.76	2010 CLIA Beckman 619 2.62-3.84 0.61-1.06   ia 2020 CLIA Beckman 228 0.72-4.76 0.58-4.66

The lower limit of FT3 (0.72 pg/mL) was found to be very low when compared with other studies. Studies suggest that lower FT3 levels and hypothyroidism are observed in fluorosis endemic area [26-28]. Flouride has multiple mechanism to interfere the thyroid function including blocking the peripheral conversion of T4 to T3 [29]. Alappuzha district, where the study was undertaken, is a well-known fluoride endemic area in Kerala [30]. Iodine nutritional status and thyroid autoimmunity may be another possible explanation [31,32]. The present study did not observe any change in FT3, FT4 and TSH levels either across different age group and or among males and females. This may be explained by smaller number of samples in each strata. The same finding was reported by Simbita M et al., from Gujrat [19]. On the contrary, the study conducted by Tannu K et al., from Ranchi reveals variation in all the three hormones according to age and sex [13]. On the contrary, Marwaha RK et al., reported no age related changes for FT3 and FT4 levels and no age and gender related changes for TSH based on a large community based study in Delhi with more than 4000 subjects recruited after stringent exclusion criteria [11]. In this study, a comparatively wider range and lower value for the lower limit of RI was noticed for FT3, FT4 and TSH and no age and gender based differences were observed.

#### Limitation(s)

Sonologic evaluation of the thyroid gland was not done in the study population. Iodine status of the population also was not assessed. The possible interference by fluoride on thyroid function could have been ruled out by assessing serum fluoride level, since Alappuzha is a flourosis endemic area. Age and sex-specific RI could not be established due to inadequate sample size for each age strata.

# CONCLUSION(S)

The present study was community based and established a RI for thyroid hormone in the local reference population selected based on the recommendation by the National Health and Nutrition Examination Survey. The RI established is found to be different from that reported by the manufacturer and studies reported from other part of the country. A large population based multicentric study should be done to establish age and gender specific RI in the state.

## REFERENCES

- [1] Kochupillai N. Clinical Endocrinology in India. 2 Current Science. 2000;8:1061-67.
- [2] Ambika G, Unnikrishnan, Usha VM. Thyroid disorders in India: An epidemiological perspective. Ind J of End and Met. 2011;15(2):78-81.
- [3] Shashank RJ. Laboratory evaluation of thyroid function. Supplement To Japi. 2011;59:14-20.

- [4] Zubair B, Pierre C, Bernard C, Laurence MD, Ulla F, Jean FH, et al. Laboratory Medicine Practice guidelines: Laboratory support for the diagnosis and monitoring of thyroid disease. Thyroid. 2003;13:03-126.
- [5] Solberg HE. The concept of reference values. J Clin Chem Clin Biochem. 1987;25:645-56.
- [6] Marwaha RK, Tandon N, Desai A, Kanwar R, Mani K. Iodine nutrition in upper socioeconomic school children of Delhi. Indian Pediatr. 2010;47:335-38.

[7] Brahmbhatt SR, Fernley R, Brahmbhatt RM, Eastman CJB. Study of biochemical prevalence of indicators for the assessment of iodine deficiency disorders in adults at field conditions at Gujarat (India). Asia Pac J Clin Nutr. 2001;10:51-57.

- [8] Henny J. IFCC recommendations for determining reference intervals. J Lab Med. 2009;33(2):45-51.
- [9] Brabant G, Beck-Peccoz P, Jarzab B, Laurberg P, Orgiazzi J, Szabolcs I, et al. Is there a need to redefine the upper normal limit of TSH? Eur J Endocrinol. 2006;154:633-37.
- [10] Laurberg P, Andersen S, Carlé A, Karmisholt J, Knudsen N, Pedersen IB. The TSH upper reference limit: Where are we at? Nat Rev Endocrinol. 2011;7:232-39.
- [11] Marwaha RK, Nikhil T, Mohd AG, Neena M, Aparna S, et al. Reference range of thyroid function (FT3, FT4 and TSH) among Indian adults. Clin Biochem. 2013;46:431-46.
- [12] Arora RK, Bara MP, Anil KK, Archana, Pushplata, Kumar A. Study of thyroid profile in advancing age. Int J of Cont Med. 2017,5(4):1169-71.
- [13] Tannu K, Anupa P, Sinha KK, Meetu L, Bharti G, Kumar S. Age and sex specific thyroid hormone profile in euthyroid subjects. Biochem Tech. 2015;6(3):1008-12.
- [14] Edward AS, Basil TD, Gregory MW, Paul D, John HE, Susan AE, et al. How to define and determine reference intervals in the Clinical Laboratory; Approved Guideline- 2<sup>nd</sup> ed. Wayne, Pa: Clinical and Laboratory Standards Institute; 2000. NCCLS document C28-A2.
- [15] Hollowell JG, Staeling NW, Flanders WD, Hannon WH, Gunter EW, Spencer CA. Serum TSH, T4, and thyroid antibodies in the United States population (1986 to 1994): National Health and Nutrition Examination Survey (NHANES III). J Clin Endocrinol Metab. 2002;87:489-99.
- [16] Gary LH, Sousan A, James CB, Ferrucio C, Uttam G, Paul H, et al. Defining, Establishing, and Verifying Reference Intervals in the Clinical Laboratory, Approved Guideline, Clinical Laboratory Standards Institute: ed 3. Villanova, PA, 2008.
- [17] Demers LM, Spencer CA, eds. Laboratory support for the diagnosis and monitoring of thyroid disease. Washington DC: National Academy Of Clinical Biochemistry, laboratory medicine practice guidelines; 2002.
- [18] Hoaglin DC, Iglewicz B, Tukey JW. Performance of some resistant rules for outlier labeling. J Am Stat Assoc. 1986;81:991-99.
- [19] Simbita M, Mihir M, Charmi R, Amit T, Haridas N. Establishment of reference intervals of thyroid function test in healthy individuals. International J Clin Biochem and Research. 2018;5(2):257-62.
- [20] Jing C, Yujie F, Jing D, Shaoyong X, Jie M, Bin G, et al. Reference intervals of thyroid hormones in a previously iodine-deficient but presently more than adequate area of Western China: A population-based survey. Endocrine J. 2016;63(4):381-88.
- [21] Henry V, Dietrich A, Thomas K, Jan Ln, Matthias N, Ulrich J, et al. Reference intervals of serum thyroid function tests in a previously iodine-deficient area. Thyroid. 2005;15(3):280-85.
- [22] Imad RM, Nagi IA, Sittan AE, Osman EO, Ishag A. Reference intervals of thyroid hormones in Khartoum, Sudan. BMC Res Notes. 2018;11:729-35.
- [23] Bosa MA, Sanja A, Tanja SJ, Darja S, Mira P, Natasa BS, et al. Direct estimation of reference intervals for thyroid parameters in the Republic of Srpska. J Med Biochem. 2017;36:137-44.
- [24] Raana A, Hafiz GA, Abubaker S, Sohail C, Sara N. Reference intervals for Free T3 and Free T4 in Pakistani euthyroid patients: Effect of age and gender on thyroid function. Journal of the College of Physicians and Surgeons Pakistan. 2014;24(11):806-09.

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- [25] Aydan CC, Sibel B, Omur E, Huriye A Baysal K. Thyroid hormone reference intervals and the prevalence of thyroid antibodies. Turk J Med Sci. 2010;40(4):665-72.
- [26] Sinhamahapatra P, Banerjee S, Lahiri S. Fluorosis and its impact on thyroid hormones: A cross-sectional study in Bankura district, West Bengal, India. Int J Res Med Sci. 2019;7:2204-09.
- [27] Nallan CC, Karunakar P, Neeharika SJ, Hima PM, Alekhya B, Nauseen S. A systematic analysis on possibility of water fluoridation causing hypothyroidism. Ind J of Dental Res. 2018;29(3):358-63.
- [28] Sachadeva S, Ahmed J, Singh B. Thyroid dysfunction associated with excess fluoride intakes: Scope for primary prevention. Thyroid Res Prc. 2015;12:50-56.
- [29] Susheela AK, Bhatnagar M, Vig K, Mondal NK. Excess fluoride ingestion and thyroid hormone derangements in children living in Delhi, India. Fluoride. 2005;38:98-108.
- [30] Gopalakrishnan P, Vasan RS, Sarma PS, Nair KS, Thankappan KR. Prevalence of dental fluorosis and associated risk factors in Alappuzha district, Kerala. Natl Med J India. 1999;12(3):99-103.
- [31] Christine DT, Jennifer M Cl, Jody M, Sheila AS. Minimal impact of excess iodate intake on thyroid hormones and selenium status in older New Zealanders. European Journal of Endocrin. 2011;165:745-52.
- [32] Hye RC. lodine and thyroid function. Ann Pediatr Endocrinol Metab. 2014;19:08-12.

#### PARTICULARS OF CONTRIBUTORS:

- 1. Professor, Department of Biochemistry, Government Thirumala Devaswom Medical College (TDMC), Alappuzha, Kerala, India.
- 2. Associate Professor, Department of Biochemistry, Government Thirumala Devaswom Medical College (TDMC), Alappuzha, Kerala, India.
- 3. Assistant Professor, Department of Biochemistry, Government Thirumala Devaswom Medical College (TDMC), Alappuzha, Kerala, India.
- 4. Biochemist, Department of Biochemistry, Government Thirumala Devaswom Medical College (TDMC), Alappuzha, Kerala, India.

#### NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR:

Gilsa Engoor Sathiandranathan,

Associate Professor, Department of Biochemistry, Government Thirumala Devaswom Medical College (TDMC), Alappuzha, Kerala, India. E-mail: mailgilsaes@gmail.com

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